

## REVIEW

# Obesity-induced microRNAs and type 2 diabetes

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Received: December 05, 2014

Published: June 27, 2015

Obesity is a growing health problem and causally linked to the development of type 2 diabetes mellitus (T2DM). T2DM is a metabolic disorder caused by peripheral insulin resistance as well as suboptimal insulin production. Although endogenous non-coding small microRNAs (miRNAs) have been shown to play an important role in the post-transcriptional repression of target genes, the implication of obesity-induced miRNAs in metabolic disorders particularly in the development of insulin resistance is largely unknown. Recent studies have revealed that saturated fatty acid (SFA) palmitate and high fat diet (HFD) significantly increase the expression of certain miRNAs, such as miR-29a and miR-195, in myocytes and hepatocytes. These obesity-induced miRNAs are also up-regulated by SFA treatment in cultured myocytes and hepatocytes. MiR-29a targets IRS-1 3'UTR directly and down-regulates IRS-1 expression at the post-transcriptional level. The induction of miR-29a impairs insulin signaling and the insulin-stimulated glucose uptake in myocytes. MiR-195 down-regulates the mRNA and protein expression of INSR without apparently changing IRS-1 expression in hepatocytes. The ectopic expression of miR-195 reduced significantly the insulin-stimulated phosphorylation of INSR and its downstream molecules in hepatocytes. Moreover, the ectopic expression of miR-195 reduced the insulin-stimulated glycogen synthesis. Taken together, these results clearly suggest that the induction of miR-29a and miR-195 by SFA are causally linked to the pathogenesis of insulin resistance and T2DM in obesity. These evidences suggest novel molecular mechanisms of insulin resistance, and implicate SFA-induced miRNAs in T2DM and metabolic diseases.

**Keywords:** microRNA; miR-29a; miR-195; palmitate, insulin resistance; IRS-1; saturated fatty acid

**To cite this article:** Won-Mo Yang, et al. Obesity-induced microRNAs in insulin resistance and type 2 diabetes. RNA Dis 2015; 2: e413. doi: 10.14800/rd.413.

## Introduction

MicroRNAs (miRNAs) are short endogenous non-coding RNAs with high evolutionary sequence conservation that regulate gene expression at post-transcriptional level [1, 2]. Mature miRNAs bind to specific complementary sequences within the 3' untranslated regions (3'UTRs) of the target genes, thereby eventually triggering either the suppression of translation or degradation of the target mRNAs [1, 2]. Although the molecular targets and roles of individual miRNAs are largely unknown, there is growing evidence suggesting that deregulated expression of miRNAs is closely

linked to various pathological states, such as cancer, neurodegeneration and cardiovascular diseases [1, 3]. Since the first discovery that miRNA is involved actively in metabolism, such as amino acid catabolism [4], several recent publications have suggested that miRNAs play significant roles in metabolic homeostasis and are implicated in the pathogenesis of insulin resistance and type 2 diabetes mellitus (T2DM) [3, 5].

Obesity is a growing public health problem worldwide, and the accumulation of excess lipid in non-adipose tissues causes lipotoxicity, which is known as dysregulation of the

cellular metabolism and apoptosis<sup>[6,7]</sup>. In particular, the accumulation of excess saturated fatty acid (SFA) in the skeletal muscle and liver has been linked to the pathogenesis of insulin resistance<sup>[8,9]</sup>. Insulin resistance is a state of reduced cellular responsiveness to insulin, and plays an important role in the development of T2DM, hypertension and metabolic diseases<sup>[8,9]</sup>. In the skeletal muscle and liver, the excess SFA results in the accumulation of diacylglycerol and provoking oxidative stress and mitochondrial dysfunction, leading to dysregulation of the cellular metabolism and insulin resistance<sup>[9-11]</sup>.

Recently, it was hypothesized that the expression of certain miRNAs targeting 3'UTR of the insulin signaling intermediates' mRNAs is modulated by the SFA-induced obesity, which participate actively in the development of insulin resistance<sup>[3,5]</sup>. Moreover, the plasma or tissue levels of those miRNAs might be associated with the insulin sensitivity of humans, thereby allowing the development of miRNAs as potential new diagnostic and therapeutic targets in insulin resistance and T2DM<sup>[3,5]</sup>. Previous our studies unveiled that the certain miRNAs, such as miR-29a and miR-195, were increased in the skeletal muscle and liver of high fat diet (HFD)-fed mice with insulin resistance<sup>[12,13]</sup>. Hence, we investigated whether these miRNAs could be induced by SFA, and whether these miRNAs might actively participate in SFA-induced insulin resistance.

MiR-29a, which is located in human chromosome 7q32.3<sup>[14]</sup>, is a member of the miR-29 family comprised of three mature forms, miR-29a, -29b and -29c<sup>[15]</sup>. Thus far, approximately 50 target genes of the miR-29 family members have been validated experimentally from thousands of predicted target genes<sup>[15]</sup>. However, it is unclear if and how miR-29a is linked to the regulation of insulin sensitivity in the skeletal muscle. We have found that the expression of miR-29a was up-regulated in the skeletal muscle, liver and serum of HFD-fed mice (60% kcal from fat for 14 weeks)<sup>[12]</sup>. In addition, the cellular treatment of SFA palmitate in myocytes increased the level of miR-29a expression<sup>[12]</sup>. *In silico* target analysis indicates that the insulin receptor substrate-1 (IRS-1) 3'UTR contains the two putative binding sites for miR-29a and is highly conserved in various species. According to the reporter gene assay, the luciferase activities of IRS-1 3'UTR containing the miR-29a binding site were reduced significantly by miR-29a, whereas the luciferase activities of IRS-1 3'UTR containing miR-29a binding site mutants were unaffected. Moreover, the ectopic expression of miR-29a reduced IRS-1 protein expression significantly without an apparent change in the IRS-1 mRNA level in myocytes<sup>[12]</sup>. Therefore, miR-29a targets IRS-1 3'UTR directly and down-regulates IRS-1 expression at the post-transcriptional level. Moreover, the ectopic expression

of miR-29a reduced the insulin-stimulated tyrosine phosphorylation of IRS-1 and its downstream molecules, such as Akt2 and glycogen synthase kinase-3 $\beta$  (GSK3 $\beta$ ) in myocytes<sup>[12]</sup>. In addition, the ectopic expression of miR-29a reduced the insulin-stimulated glucose uptake in myocytes<sup>[12]</sup>. Overall, the ectopic expression of miR-29a impairs insulin signaling and the insulin-stimulated glucose uptake in skeletal muscle cells through the post-transcriptional repression of IRS-1.

The mechanisms of SFA-induced and HFD-induced miR-29a expression in myocytes are unclear. Nevertheless, several reports have suggested that the expression of miR-29a induced several transcription factors, such as NF- $\kappa$ B, c-Myc, CEBPA, and p53 through binding the miR-29a gene promoter regions<sup>[15-18]</sup>. These transcription factors are up-regulated in HFD-fed mice<sup>[15-18]</sup>. Therefore, the up-regulation of these transcription factors by SFA and HFD might increase the expression of miR-29a. It is suggested that obesity induced by SFA generates the signals related to oxidative stress, inflammation and ER stress, and activates several transcriptional factors, such as p53 and CEBPA. These changes might up-regulate the expression of miR-29a, leading to the post-transcriptional repression of IRS-1 and subsequent insulin resistance.

Hsa-miR-195 is an intronic miRNA of a hypothetical protein, LOC284112, located on chromosome 17p13.1<sup>[19]</sup>. Based on the expression profiling and target validation analysis, the deregulation of miR-195 expression is associated with multiple diseases, such as cancer, cardiac hypertrophy and heart failure<sup>[20]</sup>. An increase of miR-195 expression by SFA also causes apoptosis via targeting Sirt1 and Bcl-2 in cardiomyocytes<sup>[21]</sup>. The other report suggests that miR-195 expression was increased in the liver of a T2DM animal model, Goto-Kakizaki rat<sup>[22]</sup>. Previously, we have found that HFD increased the miR-195 expression in the liver with a concomitant decrease in insulin receptor (INSR) and IRS-1 protein expression, suggesting that miR-195 participates in the development of hepatic insulin resistance in diet-induced obesity (DIO) mice<sup>[13]</sup>. Moreover, miR-195 expression was also found to be up-regulated significantly in palmitate-treated hepatocytes<sup>[13]</sup>. According to TargetScan and PicTar analysis, the 3'UTRs of the INSR and IRS-1 genes contain the predicted conserved binding sites for miR-195. Based on the dual luciferase reporter gene assay, the ectopic expression of miR-195 showed a significant decrease in INSR mRNA concomitant with the substantial suppression of the INSR protein in HepG2 cells<sup>[13]</sup>. On the other hand, the ectopic expression of miR-195 did not affect the mRNA and protein levels of the insulin signaling molecules, such as IRS-1, Akt2 and GSK3 $\beta$  in HepG2 cells, indicating that miR-195 down-regulates the

mRNA and protein expression of INSR without apparently changing IRS-1 expression in hepatocytes<sup>[13]</sup>. Furthermore, the ectopic expression of anti-miR-195, an antagomiR of miR-195, partially but significantly rescued the palmitate-induced suppression of INSR in HepG2 cells, indicating that the up-regulation of miR-195 contributes to palmitate-induced INSR suppression in hepatocytes<sup>[13]</sup>. Previous study also examined whether the up-regulation of miR-195 impairs insulin signaling in hepatocytes. The ectopic expression of miR-195 impairs the insulin-stimulated phosphorylation of INSR and its downstream molecules in hepatocytes<sup>[13]</sup>. Moreover, the ectopic expression of miR-195 decreased the glycogen synthesis stimulated by insulin<sup>[13]</sup>. These results clearly show that the up-regulation of miR-195 impairs insulin signaling and the insulin-stimulated glycogen synthesis in hepatocytes through post-transcriptional repression of INSR.

The mechanisms of SFA-induced and HFD-induced miR-195 expression in hepatocytes are unclear. *In silico* analysis, however, suggested that several transcription factors, such as SREBPs, PPARs, CEBPA and p53, might bind to the specific sites on the miR-195 gene promoter regions. Because the activation of SREBPs are essential for diabetic hepatic steatosis and carbohydrate-induced hypertriglyceridemia<sup>[23]</sup>, the miR-195 level in the liver may be up-regulated by SFA or HFD through SREBPs activation. PPARs are also well-known transcription factors activated by HFD that have been implicated strongly in T2DM and metabolic syndrome<sup>[24]</sup>. Another adipogenic transcription factor, CEBPA, is activated by high-energy nutrients<sup>[25]</sup>. Furthermore, the levels of CEBPA and p53 expression are up-regulated in DIO mice<sup>[18, 25]</sup>. Although further study will be needed, the induction of miR-195 in hepatocytes might be triggered by the activation of certain transcription factors regulated by SFA or DIO.

Over the last decade, studies focusing on the role of obesity-induced miRNAs in metabolism have suggested the strong prognostic and therapeutic importance of miRNAs in insulin resistance and T2DM. Previous studies have implicated obesity-induced miRNAs, such as miR-29a and miR-195, with the development of insulin resistance and T2DM. Because miR-29a and miR-195 can target several genes and each gene can be regulated by several other metabolic regulators, we are currently investigating the regulation mechanism in these miRNAs and their crosstalk with metabolic regulators. Such studies will help better understand the pathogenesis of obesity-induced insulin resistance and T2DM, and enable the development of obesity-induced miRNAs as novel diagnostic and therapeutic targets for diabetes. In addition, expression profiling of miR-29a and miR-195 in body fluids, such as plasma and

urine, will facilitate the development of new diagnostic methods by differentiating between normal and diabetic characteristics. Moreover, obesity-induced miRNAs might even find applications in therapeutics as either single agents or in combination therapies.

## ACKNOWLEDGEMENT

This study was supported by Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Education (2013R1A1A2057932).

## References

1. Bartel DP. MicroRNAs: target recognition and regulatory functions. *Cell* 2009; 136:215-233.
2. Pillai RS, Bhattacharyya SN, Filipowicz W. Repression of protein synthesis by miRNAs: how many mechanisms? *Trends Cell Biol* 2007; 17:118-126.
3. Hennessy E, O'Driscoll L. Molecular medicine of microRNAs: structure, function and implications for diabetes. *Expert Rev Mol Med* 2008; 10:e24.
4. Mersey BD, Jin P, Danner DJ. Human microRNA (miR29b) expression controls the amount of branched chain alpha-ketoacid dehydrogenase complex in a cell. *Hum Mol Genet* 2005; 14:3371-3377.
5. Park SY, Jeong HJ, Yang WM, Lee W. Implications of microRNAs in the pathogenesis of diabetes. *Arch Pharm Res* 2013; 36:154-166.
6. Samuel VT, Shulman GI. Mechanisms for insulin resistance: common threads and missing links. *Cell* 2012; 148:852-871.
7. Boren J, Taskinen MR, Olofsson SO, Levin M. Ectopic lipid storage and insulin resistance: a harmful relationship. *J Intern Med* 2013; 274:25-40.
8. Kahn SE, Hull RL, Utzschneider KM. Mechanisms linking obesity to insulin resistance and type 2 diabetes. *Nature* 2006; 444:840-846.
9. Petersen KF, Shulman GI. Etiology of insulin resistance. *Am J Med* 2006; 119:S10-16.
10. Shulman GI. Cellular mechanisms of insulin resistance. *J Clin Invest* 2000; 106:171-176.
11. Capurso C, Capurso A. From excess adiposity to insulin resistance: the role of free fatty acids. *Vascul Pharmacol* 2012; 57:91-97.
12. Yang WM, Jeong HJ, Park SY, Lee W. Induction of miR-29a by saturated fatty acids impairs insulin signaling and glucose uptake through translational repression of IRS-1 in myocytes. *FEBS Lett* 2014; 588:2170-2176.
13. Yang WM, Jeong HJ, Park SY, Lee W. Saturated fatty acid-induced miR-195 impairs insulin signaling and glycogen metabolism in HepG2 cells. *FEBS Lett* 2014; 588:3939-3946.
14. Fabbri M, Garzon R, Cimmino A, Liu Z, Zanesi N, Callegari E, et al. MicroRNA-29 family reverts aberrant methylation in lung

- cancer by targeting DNA methyltransferases 3A and 3B. *Proc Natl Acad Sci U S A* 2007; 104:15805-15810.
15. Liston A, Papadopoulou AS, Danso-Abeam D, Dooley J. MicroRNA-29 in the adaptive immune system: setting the threshold. *Cell Mol Life Sci* 2012; 69:3533-3541.
  16. Kriegel AJ, Liu Y, Fang Y, Ding X, Liang M. The miR-29 family: genomics, cell biology, and relevance to renal and cardiovascular injury. *Physiol Genomics* 2012; 44:237-244.
  17. Eyholzer M, Schmid S, Wilkens L, Mueller BU, Pabst T. The tumour-suppressive miR-29a/b1 cluster is regulated by CEBPA and blocked in human AML. *Br J Cancer* 2010; 103:275-284.
  18. Derdak Z, Villegas KA, Harb R, Wu AM, Sousa A, Wands JR. Inhibition of p53 attenuates steatosis and liver injury in a mouse model of non-alcoholic fatty liver disease. *J Hepatol* 2013; 58:785-791.
  19. Flavin RJ, Smyth PC, Laios A, O'Toole SA, Barrett C, Finn SP, *et al.* Potentially important microRNA cluster on chromosome 17p13.1 in primary peritoneal carcinoma. *Mod Pathol* 2009; 22:197-205.
  20. He JF, Luo YM, Wan XH, Jiang D. Biogenesis of MiRNA-195 and its role in biogenesis, the cell cycle, and apoptosis. *J Biochem Mol Toxicol* 2011; 25:404-408.
  21. Zhu H, Yang Y, Wang Y, Li J, Schiller PW, Peng T. MicroRNA-195 promotes palmitate-induced apoptosis in cardiomyocytes by down-regulating Sirt1. *Cardiovasc Res* 2011; 92:75-84.
  22. Herrera BM, Lockstone HE, Taylor JM, Ria M, Barrett A, Collins S, *et al.* Global microRNA expression profiles in insulin target tissues in a spontaneous rat model of type 2 diabetes. *Diabetologia* 2010; 53:1099-1109.
  23. Moon YA, Liang G, Xie X, Frank-Kamenetsky M, Fitzgerald K, Koteliansky V, *et al.* The Scap/SREBP pathway is essential for developing diabetic fatty liver and carbohydrate-induced hypertriglyceridemia in animals. *Cell Metab* 2012; 15:240-246.
  24. Sadruddin S, Arora R. Resveratrol: biologic and therapeutic implications. *J Cardiometab Syndr* 2009; 4:102-106.
  25. Um MY, Moon MK, Ahn J, Youl Ha T. Coumarin attenuates hepatic steatosis by down-regulating lipogenic gene expression in mice fed a high-fat diet. *Br J Nutr* 2013; 109:1590-1597.