

MINIREVIEW

Development of a novel cytoplasmic hydroxyl radical-targeting antioxidant (TA293) that suppresses cellular senescence, inflammation, and apoptosis

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Hydroxyl radicals ($\cdot\text{OH}$) exhibit the strongest oxidation potential of any reactive oxygen species (ROS) and react non-specifically with cellular components, such as nucleic acids, lipids and proteins. While mitochondrial $\cdot\text{OH}$ incites oxidative damage resulting in mitochondrial dysfunction, the actions of cytoplasmic $\cdot\text{OH}$ remain unknown as no cytoplasmic $\cdot\text{OH}$ -specific scavenger has been identified to date. To solve this problem, we developed the cytoplasm- and mitochondrion-specific $\cdot\text{OH}$ -targeted scavengers TA293 and mitoTA293, respectively. As expected, TA293 and mitoTA293 scavenged $\cdot\text{OH}$, but not O_2^- or H_2O_2 . Notably, TA293 scavenged pyocyanin-induced cytoplasmic $\cdot\text{OH}$, but not mitochondrial radicals induced by antimycin A. Conversely, mitoTA293 scavenged $\cdot\text{OH}$ only in the mitochondria *in vivo* and *in vitro*. Interestingly, we found that cytoplasmic $\cdot\text{OH}$ plays a central role in cytoplasm ROS-induced oxidative stress, which potentiates cellular senescence, inflammation, and apoptosis in the kidney and lung. Based on these findings, we believe that TA293 could be a novel tool to study the effects of $\cdot\text{OH}$ damage within the cytoplasm.

Keywords: antioxidant; hydroxyl radicals; oxidative stress; cellular senescence; inflammation

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Introduction

Excess reactive oxygen species (ROS) or free radicals oxidize cellular components, resulting in oxidative stress [1, 2]. These molecules react non-specifically with nucleic acids, lipids, and proteins, and are critical mediators of age-related diseases such as cancer, type II diabetes, neurodegenerative disease, cardiovascular disease, macular degeneration, and osteoarthritis [3-8]. NADPH oxidase and xanthine oxidase in

the mitochondrial electron transport chain generate most of the superoxide anion radicals (O_2^-), which are subsequently converted to H_2O_2 by either glutathione peroxidase or catalase [9]. Excess H_2O_2 then results in the production of hydroxyl radicals ($\cdot\text{OH}$) via metal ion catalysis, as in the Fenton reaction [10]. In low concentrations, O_2^- and H_2O_2 act as signaling effectors to play important physiological roles in apoptosis, cell proliferation, and differentiation, amongst others [11]. However, unlike that of O_2^- and H_2O_2 , $\cdot\text{OH}$

scavenging is a critical antioxidant process, as there is no known detoxification system for these radicals^[12].

Recent reports have identified important roles for $\cdot\text{OH}$ in the pathophysiology of several diseases^[13-15]. Mitochondrial $\cdot\text{OH}$ are predicted to oxidize organelle constituents and cause mitochondrial dysfunction^[16]; however, the functional significance of cytoplasmic $\cdot\text{OH}$ remains to be determined since there is no way to scavenge these radicals specifically. Other $\cdot\text{OH}$ scavengers such as, thiourea, H_2 , salicylates, and flavonoids are likely distributed throughout the cell because of their low molecular-weight^[16-19], and scavenge $\cdot\text{OH}$ indiscriminately. In addition, many of these compounds have cytotoxic effects, such as damage of DNA, proteins, lipids, and other macromolecules^[20-22]. As such, the function of compartment-specific $\cdot\text{OH}$ are still unknown.

We recently developed TA293, a novel antioxidant based on the structure of ascorbic acid (also known as vitamin C). Ascorbic acid is a water-soluble molecule with antioxidant activity derived from its enediol structure and oxidized to dehydroascorbic acid via a radical intermediate^[23]. Similarly, TA293 was designed to act as resonance system by adding plural C-C double bonds to the enediol structure; thus, the structure becomes resonance-stabilized after delocalization of its radical, thereby suppressing the oxidation reaction. In addition, TA293 is a more lipophilic compound that can permeate various tissue^[24]. We also synthesized mitochondrial-specific mitoTA293 by adding the mitochondrial localization signal triphenylphosphonium to the TA293 molecule^[25]. A subsequent analysis of the effects of pyocyanin-induced cytoplasmic $\cdot\text{OH}$ in the presence of TA293 demonstrated the radical's central role in inflammation, cellular senescence, and apoptosis.

Effects of TA293 and mitoTA293

A preliminary analysis of TA293 and mitoTA293 function revealed that these compounds exhibit $\cdot\text{OH}$ scavenging activity, but have no effect on O_2^- and H_2O_2 in a cell-free system. To study the effect of each compound *in vivo*, cells were treated with pyocyanin and antimycin A to induce ROS formation in the cytoplasm and mitochondria, respectively^[26, 27]. As expected, TA293 appears to selectively scavenge $\cdot\text{OH}$ in the cytoplasm, whereas mitoTA293 acts solely in the mitochondria. Moreover, conventional $\cdot\text{OH}$ scavengers such as thiourea can induce cytotoxicity, but neither TA293 nor mitoTA293 hindered the viability of established cell lines or primary cells in culture. Thus, these data suggest that TA293 and mitoTA293 may help elucidate the physiological significance of $\cdot\text{OH}$ localized to the cytoplasmic or mitochondria.

TA293 scavenges pyocyanin-induced cytoplasmic $\cdot\text{OH}$.

To further elucidate the effects of TA293 and mitoTA293, we evaluated oxidative stress in primary mouse embryonic fibroblasts (MEF) derived from Keap1-dependent oxidative stress luciferase indicator transgenic (OKD48-Tg) mice that harbor an antioxidant response element (ARE)-regulated luciferase reporter^[28]. Notably, TA293 suppressed pyocyanin-induced oxidative stress and apoptosis, but not that induced by antimycin A treatment; while the opposite was true for mitoTA293. Moreover, we evaluated oxidative stress activity in pyocyanin-treated OKD48-Tg mice in the presence or absence of TA293. As expected, TA293 suppressed systemic pyocyanin-induced oxidative stress and biooxidation, but not that resulting from antimycin A. Conversely, mitoTA293 only suppressed antimycin A-induced oxidative stress and biooxidation. Together, these findings suggest that cytoplasmic and mitochondrial $\cdot\text{OH}$ play key roles in oxidative stress within their respective compartments *in vitro* and *in vivo*.

We then attempted to identify the organ most impacted by the presence of cytoplasmic $\cdot\text{OH}$. This analysis revealed that pyocyanin induced-oxidative stress is markedly increased in the lung and kidney, but not other organs such as the heart, liver, stomach, and spleen. Significantly, TA293 treatment attenuated oxidative stress in these areas, supporting that cytoplasmic $\cdot\text{OH}$ specifically induced oxidative stress in lung and kidney *in vivo*. Subsequent histological examination showed pyocyanin-induced oxidative damage in the lung alveoli and kidney tubules, which was mitigated in TA293-treated counterparts. Collectively, these data indicate that the cytoplasmic $\cdot\text{OH}$ play a central role in oxidative damage that occurs in organs highly sensitive to radicals in this subcellular compartment.

TA293 suppresses cytoplasmic $\cdot\text{OH}$ -induced cellular senescence, inflammation, and apoptosis.

Recent studies indicate that pyocyanin elicits oxidative stress and premature cellular senescence^[29]. Senescent cells secrete senescence-associated secretory phenotype (SASP) factor, such as IL-6, resulting in chronic inflammation, macrophage infiltration, senescent cell clearance, and tissue remodeling^[30, 31]. Notably, we found that TA293 suppressed cellular senescence induced by pyocyanin *in vitro*. The CDK inhibitors p21 (WAF1/ CIP1) and p16 (INK4A) induce cell cycle arrest and cellular senescence^[32], and are highly expressed in response to oxidative damaged tissues; however, TA293 dampened the expression of these factors.

I κ B phosphorylation-mediated NF- κ B signaling has also been shown to promote SASP factor secretion^[33].

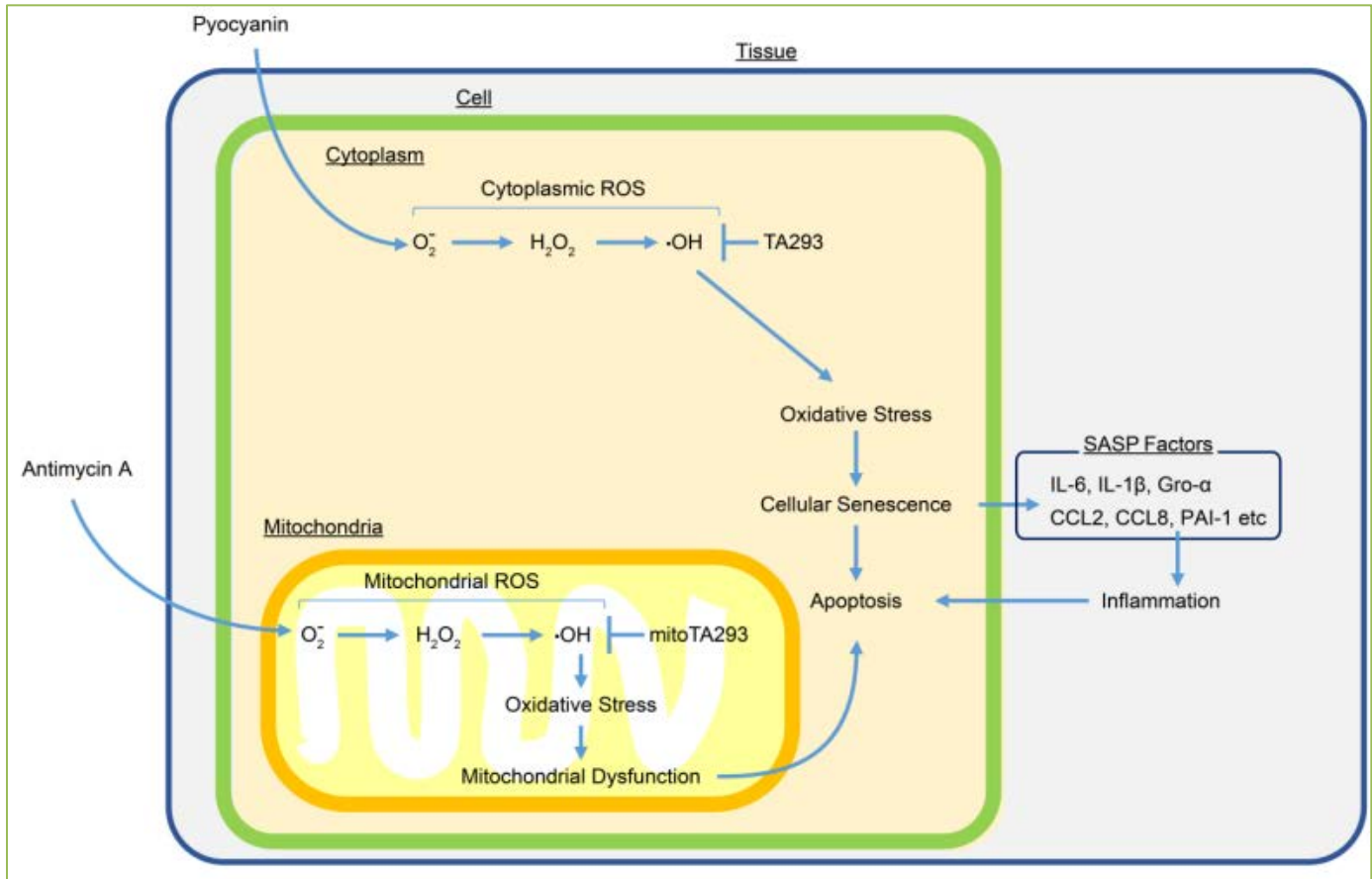


Figure 1. Cytoplasmic $\cdot\text{OH}$ -induced cellular senescence, inflammation, and apoptosis. TA293 attenuated cytoplasmic $\cdot\text{OH}$ -induced oxidative stress, cellular senescence, inflammation, and apoptosis; whereas mitoTA293 suppressed mitochondrial $\cdot\text{OH}$ -induced oxidative stress and apoptosis. These findings suggested that cytoplasmic and mitochondrial $\cdot\text{OH}$ induce oxidative damage in their respective subcellular compartments.

Consistently, we found elevated I κ B phosphorylation in tissues with oxidative damage, but this was attenuated in the presence of TA293. In addition, TA293 was sufficient to block pyocyanin-induced inflammation in IL-1 β -based dual-operating luciferase transgenic (IDOL-Tg) mice^[34]. Moreover, TA293 suppressed mRNA expression of SASP factors (e.g., proinflammatory cytokines, chemokines, and extracellular matrix remodeling factor) and apoptosis in tissues with oxidative damage in pyocyanin-treated mice. Since cellular senescence, inflammation, and apoptosis are triggered by common stimuli such as DNA damage or oncogenic stress^[35], it is not surprising that cytoplasmic $\cdot\text{OH}$ -induced oxidative damage elicited these cellular responses *in vivo* and could be attenuated by TA293 treatment.

Conclusions

In summary, the results of our study provide scientific evidence that TA293 suppressed cytoplasmic $\cdot\text{OH}$ -induced oxidative stress, cellular senescence, inflammation, and

apoptosis; whereas mitoTA293 attenuated mitochondrial $\cdot\text{OH}$ -induced oxidative stress and apoptosis (Fig. 1). Based on these findings, we propose a novel hypothesis that cytoplasmic and mitochondrial $\cdot\text{OH}$ respectively induce oxidative damage in cytoplasm and mitochondria, resulting in oxidative stress in each individual subcellular compartment.

Most importantly, this study provided the first experimental evidence that TA293 is a non-toxic compound capable of specifically scavenging cytoplasmic $\cdot\text{OH}$. For this reason, we believe that TA293 will be a useful tool to elucidate the mechanisms of $\cdot\text{OH}$ -induced inflammation, cellular senescence, and apoptosis; and may serve as a prototype to develop therapeutic agents for cytoplasmic $\cdot\text{OH}$ -induced disease.

Conflicting interests

The authors have declared that no conflict of interests exist.

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Author contributions

TS made substantial contributions to conception and design, or acquisition of data, or analysis and interpretation of data; has been involved in drafting the manuscript or revising it critically for important intellectual content; and has given final approval of the version to be published. JI, TI, HT, MU, and SH were in charge of the study design and also approved the version to be published.

Abbreviations

[•]OH: hydroxyl radical; ROS: reactive oxygen species; MEF: mouse embryonic fibroblast; Tg: transgenic; SASP: senescence-associated secretory phenotype.

References

- Wallace DC. A mitochondrial paradigm of metabolic and degenerative diseases, aging, and cancer: a dawn for evolutionary medicine. *Annu Rev Genet* 2005; 39:359-407.
- Wright E Jr, Scism-Bacon JL, Glass LC. Oxidative stress in type 2 diabetes: the role of fasting and postprandial glycaemia. *Int J Clin Pract* 2006; 60:308-314.
- Ames BN. Dietary carcinogens and anticarcinogens. Oxygen radicals and degenerative diseases. *Science* 1983; 4617:1256-1264.
- Kaneto H, Katakami N, Matsuhisa M, Matsuoka TA. Role of reactive oxygen species in the progression of type 2 diabetes and atherosclerosis. *Mediators Inflamm* 2010; 2010:453892.
- Andersen JK. Oxidative stress in neurodegeneration: cause or consequence? *Nat Med* 2004; 10:18-25.
- Shah AM, Channon KM. Free radicals and redox signalling in cardiovascular disease. *Heart* 2004; 90:486-487.
- Jarrett SG, Boulton ME. Consequences of oxidative stress in age-related macular degeneration. *Mol Aspects Med* 2012; 33:399-417.
- Lepetsos P, Papavassiliou AG. ROS/oxidative stress signaling in osteoarthritis. *Biochim Biophys Acta* 2016; 1862:576-591.
- Chinopoulos C, Adam-Vizi V. Calcium, mitochondria and oxidative stress in neuronal pathology. Novel aspects of an enduring theme. *FEBS J* 2006; 273:433-450.
- Halliwell B, Gutteridge JM. Biologically relevant metal ion-dependent hydroxyl radical generation. An update. *FEBS Lett* 1992; 307:108-112.
- Liu H, Colavitti R, Rovira II, Finkel T. Redox-dependent transcriptional regulation. *Circ Res* 2005; 97:967-974.
- Sheu SS, Nauduri D, Anders MW. Targeting antioxidants to mitochondria: a new therapeutic direction. *Biochim Biophys Acta* 2006; 1762:256-265.
- Amitani H, Asakawa A, Cheng K, Amitani M, Kaimoto K, Nakano M, et al. Hydrogen improves glycemic control in type 1 diabetic animal model by promoting glucose uptake into skeletal muscle. *PLoS One* 2013; 8:e53913.
- Zhao L, Zhou C, Zhang J, Gao F, Li B, Chuai Y, et al. Hydrogen protects mice from radiation induced thymic lymphoma in BALB/c mice. *Int J Biol Sci* 2011; 7:297-300.
- He J, Xiong S, Zhang J, Wang J, Sun A, Mei X, et al. Protective effects of hydrogen-rich saline on ulcerative colitis rat model. *J Surg Res* 2013; 185:174-181.
- Ohsawa I, Ishikawa M, Takahashi K, Watanabe M, Nishimaki K, Yamagata K, et al. Hydrogen acts as a therapeutic antioxidant by selectively reducing cytotoxic oxygen radicals. *Nat Med* 2007; 13:688-694.
- Araújo MC, Antunes LM, Takahashi CS. Protective effect of thiourea, a hydroxyl-radical scavenger, on curcumin-induced chromosomal aberrations in an in vitro mammalian cell system. *Teratog Carcinog Mutagen* 2001; 21:175-180.
- Althaus JS, Andrus PK, Williams CM, VonVoigtlander PF, Cazars AR, Hall ED. The use of salicylate hydroxylation to detect hydroxyl radical generation in ischemic and traumatic brain injury. *Mol Chem Neuropathol* 1993; 20:147-162.
- Chen JW, Zhu ZQ, Hu TX, Zhu DY. Structure-activity relationship of natural flavonoids in hydroxyl radical-scavenging effects. *Acta Pharmacol Sin* 2002; 23:667-672.
- Cohen G, Heikkila RE, Allis B, Cabbat F, Dembiec D, MacNamee D, et al. Destruction of sympathetic nerve terminals by 6-hydroxydopamine: protection by 1-phenyl-3-(2-thiazolyl)-2-thiourea, diethylthiocarbamate, methimazole, cysteamine, ethanol and n-butanol. *J Pharmacol Exp Ther* 1976; 199:336-352.
- Bellosillo B, Piqué M, Barragán M, Castaño E, Villamor N, Colomer D, et al. Aspirin and salicylate induce apoptosis and activation of caspases in B-cell chronic lymphocytic leukemia cells. *Blood* 1998; 92:1406-1414.
- Matsuo M, Sasaki N, Saga K, Kaneko T. Cytotoxicity of flavonoids toward cultured normal human cells. *Biol Pharm Bull* 2005; 28:253-259.
- Du J, Cullen JJ, Buettner GR. Ascorbic acid: chemistry, biology and the treatment of cancer. *Biochim Biophys Acta* 2012; 1826:443-457.
- Sakai T, Imai J, Ito T, Takagaki H, Ui M, Hata S. The novel antioxidant TA293 reveals the role of cytoplasmic hydroxyl radicals in oxidative stress-induced senescence and inflammation. *Biochem Biophys Res Commun* 2017; 482:1183-1189.
- Smith RA, Murphy MP. Animal and human studies with the mitochondria-targeted antioxidant MitoQ. *Ann N Y Acad Sci* 2010; 1201:96-103.
- Rada B, Jendrysik MA, Pang L, Hayes CP, Yoo DG, Park JJ, et al.

- Pyocyanin-enhanced neutrophil extracellular trap formation requires the NADPH oxidase. *PLoS One* 2013; 8:e54205.
27. Balaban RS, Nemoto S, Finkel T. Mitochondria, oxidants, and aging. *Cell* 2005; 120:483-495.
 28. Oikawa D, Akai R, Tokuda M, Iwawaki T. A transgenic mouse model for monitoring oxidative stress. *Sci Rep* 2012; 2:229.
 29. Muller M. Premature cellular senescence induced by pyocyanin, a redox-active *Pseudomonas aeruginosa* toxin. *Free Radic Biol Med* 2006; 41:1670-1677.
 30. Wynn TA, Ramalingam TR. Mechanisms of fibrosis: therapeutic translation for fibrotic disease. *Nat Med* 2012; 18:1028-1040.
 31. Muñoz-Espín D, Cañamero M, Maraver A, Gómez-López G, Contreras J, Murillo-Cuesta S, *et al.* Programmed cell senescence during mammalian embryonic development. *Cell* 2013; 155:1104-1118.
 32. Ohtani N, Hara E. Roles and mechanisms of cellular senescence in regulation of tissue homeostasis. *Cancer Sci* 2013; 104:525-530.
 33. Salminen A, Kauppinen A, Kaarniranta K. Emerging role of NF- κ B signaling in the induction of senescence-associated secretory phenotype (SASP). *Cell Signal* 2012; 24:835-845.
 34. Iwawaki T, Akai R, Oikawa D, Toyoshima T, Yoshino M, Suzuki M, *et al.* Transgenic mouse model for imaging of interleukin-1 β -related inflammation in vivo. *Sci Rep* 2015; 24:17205.
 35. Rodier F, Campisi J. Four faces of cellular senescence. Four faces of cellular senescence. *J Cell Biol* 2011; 192:547-556.