REVIEW

The functions and signaling pathways induced by the interactions of APP–related molecules with p75NTR

Yuki Fujita^{1, 2}, Toshihide Yamashita^{1, 2}

¹Department of Molecular Neuroscience, Graduate School of Medicine, Osaka University, 2-2 Yamadaoka, Suita, Osaka 565- 0871, Japan 2 JST, CREST, 5, Sanbancho, Chiyoda-ku, Tokyo, Japan

Correspondence: Toshihide Yamashita E-mail: yamashita@molneu.med.osaka-u.ac.jp Received: August 03, 2014 Published online: September 22, 2014

> **p75 neurotrophin receptor (p75NTR) regulates diverse functions, including survival, differentiation, growth, and apoptosis of neurons, through its association with a number of molecules. Accumulating evidence shows that βamyloid precursor protein (APP)–related molecules, which also regulate multiple neuronal functions, interact with p75NTR. APP is cleaved by secretases to generate several proteins including soluble β-amyloid precursor protein alpha (sAPPα), sAPPβ, and amyloid β (Aβ). Binding of Aβ to p75NTR induces neuronal death. In contrast, sAPPα directly interacts with p75NTR to enhance neurite outgrowth through the activation of protein kinase A (PKA). This review focuses on the molecular mechanisms and functions occurring as a consequence of interactions of p75NTR with APP–related molecules.**

Keywords: APP; sAPP alpha; p75^{NTR}; neurite outgrowth; central nervous system

To cite this article: Yuki Fujita, *et al*. The functions and signaling pathways induced by the interactions of APP–related molecules with p75^{NTR}. Receptor Clin Invest 2014; 1: e283. doi: 10.14800/rci.283.

Copyright: © 2014 The Authors. Licensed under a *Creative Commons Attribution 4.0 International License* which allows users including authors of articles to copy and redistribute the material in any medium or format, in addition to remix, transform, and build upon the material for any purpose, even commercially, as long as the author and original source are properly cited or credited.

Introduction

p75NTR is a member of a large family of receptors, including tumor necrosis factor (TNF) receptors, Fas (Apo-1/CD95), RANK, CD40, and approximately 25 other members. p75NTR mediates diverse functions, such as axonal elongation, neuronal survival, and modulation of synaptic transmission. $p75^{NTR}$ regulates both positive and negative signals for certain neuronal functions $[1, 2]$. These bi-directional signals can be explained by the complex formation of p75NTR with other receptors and multiple downstream intracellular signaling molecules. For example, p75^{NTR} is known to regulate axonal elongation

both positively and negatively. Neurotrophin binding to p75NTR decreases the activation of Rho small GTPase, resulting in the promotion of axonal elongation^[3]. Indeed, p75NTR-deficient mice show retarded outgrowth of intercostal nerves in the developmental stage [3]. In contrast, p75NTR is also involved in axon growth inhibition through the activation of RhoA in the presence of myelin– derived inhibitors, such as myelin–associated glycoprotein (MAG), Nogo, and oligodendrocyte glycoprotein (OMgp) $^{[4]}$. In this case, p 75^{NTR} interacts with the Nogo receptor to form a receptor complex for these proteins ^[5]. Thus, the binding partners of p75^{NTR} may determine its downstream signaling and function.

p75^{NTR} associates with APP–related molecules ^[6-8]. Aβ peptide that accumulates in Alzheimer's disease (AD) induces cell death via $p75^{NTR}$ in multiple types of cells, including some types of neurons $[8-11]$. A recent study demonstrated that the N-terminal fragment of APP (N-APP) interacts with $p75^{NTR}$ [12]. A β and APP often show neurotoxicity, whereas sAPPα, which is a product of the non-amyloidogenic APP cleavage by α-secretase, has neuroprotective effects. Binding of sAPPα to p75NTR stimulates neurite outgrowth $[7]$. In this review, we focus on the functions and signals induced by the interaction of p75NTR with APP–related molecules in the central nervous systems (CNS).

Molecular profile of APP-related molecules

APP isoforms

APP is a single transmembrane protein with a long Nterminal domain in the extracellular space and a short Cterminal domain in the cytosol $[13]$. In mammals, the APP super family consists of APP itself, and APP-like proteins 1 and 2 (APLP1 and APLP2). APLP1 and APLP2 share sequence similarity with APP, although they lack $A\beta$ domain [14,15] . APLP2 as well as APP are expressed ubiquitously, whereas APLP1 is exclusively expressed in the brain. The APP genes are located on chromosome 21 [16]. Alternative mRNA splicing of exon 7 and 8 generates the three major APP isoforms (APP695, APP751, and APP770) in the CNS $[17-22]$. APP751 and APP770, but not APP695, contain a Kunitz protease inhibitor (KPI) domain. In the brain, under the physiological condition, APP695 is predominantly expressed. In the AD brain, the expression levels of the other two APP isoforms seem to be increased $^{[23-25]}$. These findings suggest that changes in the expression of each APP isoform may be associated with AD.

Proteolysis of APP

APP undergoes post-translational proteolysis to generate multiple fragments. These proteolytic products elicit pathological and physiological functions (Figure 1). There are two distinct pathways of APP metabolism: the non-amyloidogenic pathway, which avoids Aβ generation, and the amyloidogenic pathway, which generates Aβ. In the non-amyloidogenic pathway, APP is cleaved by αsecretase, releasing the large N-terminal ectodomain, sAPP α , from the cell surface. The remaining 83-aminoacid-long C-terminal fragment (C83) remains in the membrane. This fragment can be subsequently cleaved by γ-secretase, giving rise to small fragments, p3 and APP intracellular domain (AICD). In this pathway, the generation of Aβ is avoided, since α-secretase cleavage occurs within the Aβ sequence near the extracellular side

Figure 1. APP-related molecules and p75NTR . α-secretase catalyzes the cleavage of APP to generate a soluble peptide, sAPPα. β-secretase cleaves APP to generate an alternate soluble peptide, sAPPβ, which is cleavedby γ-secretase to generate Aβ. The interaction of p75^{NTR} with Aβ or APP leads to neuronal death. In contrast, the interaction of p75^{NTR} with sAPPα promotes neuroprotection and neurite outgrowth through the activation of PKA.

of the plasma membrane $[26, 27]$. In the amyloidogenic pathway, APP is cleaved by β-secretase, which leads to the release of the N-terminal ectodomain, sAPPβ. The remaining 99-amino-acid C-terminal stub (C99) can be further cleaved by γ-secretase in a similar way as in the non-amyloidogenic pathway. This produces AICD and the amyloidogenic Aβ peptide^[28].

Interactions of APP-related molecules with p75NTR

In the adult brain, $p75^{NTR}$ expression is restricted in the CNS, and the highest expression is observed in the neurons in the cholinergic basal forebrain, which represents a vulnerable region in AD^{$[29-32]$}. p75^{NTR} mediates apoptosis induced by pro–nerve growth factor (pro-NGF), which is increased in the brains of AD subjects [33]. These observations suggest a possible association of p75NTR and AD pathogenesis.

Although the expression levels of $p75^{NTR}$ in the brains of patients with AD are contentious, most studies conclude that AD is associated with thedownregulation of tropomyosin-related kinase A (TrkA) expression with changed/unchanged expression of p75^{NTR [34-36]}. The expression levels of $A\beta1-42$ and $p75^{NTR}$ in the

hippocampal cells from 12–15 month–old AD-triple transgenic mice (3xTg-AD mouse, which expresses APPs we, PS1M146V, and tauP301L mutations) are increased compared to that in hippocampal cells from age–matched wild-type mice ^[37]. These results are consistent with another observation that demonstrates elevated expression of p75NTR in hippocampal CA1 and CA2 neurons in human AD brains [34].

Accumulating evidence has revealed that $p75^{NTR}$ directly interacts with Aβ and APP, and that this interaction leads to apoptosis $[6, 8]$. It has been shown that aggregated Aβ induces apoptosis in NIH-3T3 cells stably expressing p75NTR, but not wild-type control cells lacking the receptor. Normal neural crest–derived melanocytes, endogenously expressing p75^{NTR}, undergo apoptosis in the presence of Aβ^[8]. Binding of Aβ to p75^{NTR} induces the transcription of c-Jun mRNA. This leads to the activation of c-Jun N-terminal kinase (JNK) [38]. These results suggest that the interaction of $p75^{NTR}$ with A β induces apoptosis through the activation of JNK $[8-10, 38]$.

Further, p75^{NTR} is also involved in neuronal damage by interacting with APP. $p75^{NTR}$ directly interacts with APP, and the co-expression of $p75^{NTR}$ and APP induces cell death $^{[6]}$. The interaction of p75^{NTR} with APP is blocked in the presence of the ligands for $p75^{NTR}$, A β or NGF. Taken together, the interaction of $p75^{NTR}$ with A β and APP contributes to neuronal vulnerability in AD.

The effects of sAPPα on the neurons related with p75NTR

Neuroprotective role of sAPPα

α-secretase catalyzes the cleavage of APP to generate sAPPα. This cleavage occurs within the Aβ sequence, avoiding Aβ generation. Unlike Aβ and APP, sAPPα has been shown to demonstrate neuroprotective effects both *in vivo* and *in vitro*. In the *in vivo* model of transient ischemia, administration of sAPPα protects CA1 hippocampal neurons against ischemic injury ^[39]. Treatment with sAPPα following a traumatic brain injury in rats improves functional outcome [40]. The growth factor-like domain (D1) and the E2 domain within the carbohydrate domain (D6a) of sAPP α (residues 28–123 and 316–498), which are able to bind to heparan sulfate proteoglycans (HSPGs), is found to be involved in the improvement of motor and cognitive outcomes against traumatic brain injury in rats, suggesting that HSPGs could mediate this response [41]. Furthermore, animal models that over express $sAPP\alpha$ have been developed. Knock-in of sAPPα rescues the abnormal phenotype of APP knockout mouse, such as reductions in the brain and body weight, grip strength deficits, and the impairment in spatial learning and LTP $[42]$. Transgenic mice with neuronal over-expression of APP, mutated at the α-secretase site, demonstrate the increased aggressiveness, disturbed responses to kainic acid and N-methyl-Daspartate (NMDA), neophobia, and deficiency in exploratory behavior $[43, 44]$. These animal models confirm that sAPPα plays beneficial roles in the neurons.

sAPPα shows neuroprotective function against Aβ peptide-induced oxidative injury and glucose deprivation *in vitro* [45,46] . The C-terminus of sAPPα (residues 591– 612) mediates *in vitro* neuroprotective effects against glutamate and Aβ toxicity, and heparinases greatly reduce this action $^{[47]}$. sAPP α also protects PC12 cells and mature neurons from other toxic insults such as epoxomicin or UV irradiation by inhibiting the stress-triggered JNK-signaling pathway^[48].

Neurite outgrowth and sAPPα

sAPP α has been shown to enhance neurite outgrowth [49,50] . Both the secreted forms of APP695 and APP770 promote neurite outgrowth in embryonic rat neocortical explants. This indicates that KPI domain is not responsible for neurite outgrowth-promoting activity of sAPPα.

The interaction of $p75^{NTR}$ with sAPP α contributes to sAPP α -induced neurite outgrowth [7]. Direct interaction of sAPP α with p75^{NTR} was identified by coimmunoprecipitation and enzyme-linked immunosorbent assay (ELISA). EC50 of the interaction between sAPPα and p75NTR is 90 nM. In contrast, the affinity of NGF for $p75^{NTR}$ is higher (K_d 1–2 nM) ^[51, 52]. These observations indicate that the binding affinity of sAPP α for p75^{NTR} is lower than that of neurotrophins. Furthermore, Binding of sAPP α to p75^{NTR} promotes neurite outgrowth in the embryonic mouse cortical neurons. Protein kinase A (PKA) inhibitor KT5720 inhibits this function, suggesting that PKA activation is essential for sAPPα-induced neurite outgrowth.

Conclusions

As reviewed here, the interaction of $p75^{NTR}$ with A β and APP promotes neuronal cell death, while the interaction of $p75^{NTR}$ with sAPP α contributes to neurite outgrowth through the activation of PKA (Figure 1). These findings suggest that the balance of α-cleavage and β-cleavage of APP is critical to control cellular susceptibility to neurotoxic insults. It is notable that p75NTR mediates these opposite cues. The balancing mechanisms through p75^{NTR} may regulate the CNS development and the determination of cellular fate after CNS injury.

Conflict of interests

The authors declare that they have no conflict of

interests.

References

- 1. Dechant G, Barde YA. The neurotrophin receptor p75(NTR): novel functions and implications for diseases of the nervous system. Nat Neurosci 2002; 5: 1131-1136.
- 2. Kaplan DR, Miller FD. Neurotrophin signal transduction in the nervous system. Curr Opin Neurobiol 2000; 10: 381-391.
- 3. Yamashita T, Tucker KL, Barde YA. Neurotrophin binding to the p75 receptor modulates Rho activity and axonal outgrowth. Neuron 1999; 24: 585-593.
- 4. Yamashita T, Higuchi H, Tohyama M. The p75 receptor transduces the signal from myelin-associated glycoprotein to Rho. J Cell Biol 2002; 157: 565-570.
- 5. Wang KC, Kim JA, Sivasankaran R, Segal R, He Z. P75 interacts with the Nogo receptor as a co-receptor for Nogo, MAG and OMgp. Nature 2002; 420: 74-78.
- 6. Fombonne J, Rabizadeh S, Banwait S, Mehlen P, Bredesen DE. Selective vulnerability in Alzheimer's disease: amyloid precursor protein and p75(NTR) interaction. Ann Neurol 2009; 65: 294-303.
- 7. Hasebe N, Fujita Y, Ueno M, Yoshimura K, Fujino Y, Yamashita T. Soluble beta-amyloid Precursor Protein Alpha Binds to p75 Neurotrophin Receptor to Promote Neurite Outgrowth. PLoS One 2013; 8: e82321.
- 8. Yaar M, Zhai S, Pilch PF, Doyle SM, Eisenhauer PB, Fine RE, *et al*. Binding of beta-amyloid to the p75 neurotrophin receptor induces apoptosis. A possible mechanism for Alzheimer's disease. J Clin Invest 1997; 100: 2333-2340.
- 9. Kuner P, Hertel C. NGF induces apoptosis in a human neuroblastoma cell line expressing the neurotrophin receptor p75NTR. J Neurosci Res 1998; 54: 465-474.
- 10. Perini G, Della-Bianca V, Politi V, Della Valle G, Dal-Pra I, Rossi F, *et al*. Role of p75 neurotrophin receptor in the neurotoxicity by beta-amyloid peptides and synergistic effect of inflammatory cytokines. J Exp Med 2002; 195: 907-918.
- 11. Sotthibundhu A, Sykes AM, Fox B, Underwood CK, Thangnipon W, Coulson EJ. Beta-amyloid(1-42) induces neuronal death through the p75 neurotrophin receptor. J Neurosci 2008; 28: 3941-3946.
- 12. Nikolaev A, McLaughlin T, O'Leary DD, Tessier-Lavigne M. APP binds DR6 to trigger axon pruning and neuron death via distinct caspases. Nature 2009; 457: 981-989.
- 13. Kang J, Lemaire HG, Unterbeck A, Salbaum JM, Masters CL, Grzeschik KH, *et al*. The precursor of Alzheimer's disease amyloid A4 protein resembles a cell-surface receptor. Nature 1987; 325: 733-736.
- 14. Sprecher CA, Grant FJ, Grimm G, O'Hara PJ, Norris F, Norris K, *et al*. Molecular cloning of the cDNA for a human amyloid precursor protein homolog: evidence for a multigene family. Biochemistry 1993; 32: 4481-4486.
- 15. Wasco W, Gurubhagavatula S, Paradis MD, Romano DM, Sisodia SS, Hyman BT, *et al*. Isolation and characterization of APLP2 encoding a homologue of the Alzheimer's associated amyloid beta protein precursor. Nat Genet 1993; 5: 95-100.
- 16. Goldgaber D, Lerman MI, McBride OW, Saffiotti U, Gajdusek DC. Characterization and chromosomal localization of a cDNA

encoding brain amyloid of Alzheimer's disease. Science 1987; 235: 877-880.

- 17. Kang J, Muller-Hill B. Differential splicing of Alzheimer's disease amyloid A4 precursor RNA in rat tissues: PreA4(695) mRNA is predominantly produced in rat and human brain. Biochem Biophys Res Commun 1990; 166: 1192-1200.
- 18. Golde TE, Estus S, Usiak M, Younkin LH, Younkin SG. Expression of beta amyloid protein precursor mRNAs: recognition of a novel alternatively spliced form and quantitation in Alzheimer's disease using PCR. Neuron 1990; 4: 253-267.
- 19. Kitaguchi N, Takahashi Y, Tokushima Y, Shiojiri S, Ito H. Novel precursor of Alzheimer's disease amyloid protein shows protease inhibitory activity. Nature 1988; 331: 530-532.
- 20. Neve RL, Finch EA, Dawes LR. Expression of the Alzheimer amyloid precursor gene transcripts in the human brain. Neuron 1988; 1: 669-677.
- 21. Ponte P, Gonzalez-DeWhitt P, Schilling J, Miller J, Hsu D, Greenberg B, *et al*. A new A4 amyloid mRNA contains a domain homologous to serine proteinase inhibitors. Nature 1988; 331: 525-527.
- 22. Tanzi RE, McClatchey AI, Lamperti ED, Villa-Komaroff L, Gusella JF, Neve RL. Protease inhibitor domain encoded by an amyloid protein precursor mRNA associated with Alzheimer's disease. Nature 1988; 331: 528-530.
- 23. Menendez-Gonzalez M, Perez-Pinera P, Martinez-Rivera M, Calatayud MT, Blazquez Menes B. APP processing and the APP-KPI domain involvement in the amyloid cascade. Neurodegener Dis 2005; 2: 277-283.
- 24. Moir RD, Lynch T, Bush AI, Whyte S, Henry A, Portbury S, *et al*. Relative increase in Alzheimer's disease of soluble forms of cerebral Abeta amyloid protein precursor containing the Kunitz protease inhibitory domain. J Biol Chem 1998; 273: 5013- 5019.
- 25. Tanaka S, Shiojiri S, Takahashi Y, Kitaguchi N, Ito H, Kameyama M, *et al*. Tissue-specific expression of three types of beta-protein precursor mRNA: enhancement of protease inhibitor-harboring types in Alzheimer's disease brain. Biochem Biophys Res Commun 1989; 165: 1406-1414.
- 26. Esch FS, Keim PS, Beattie EC, Blacher RW, Culwell AR, Oltersdorf T, *et al*. Cleavage of amyloid beta peptide during constitutive processing of its precursor. Science 1990; 248: 1122-1124.
- 27. Sisodia SS, Koo EH, Beyreuther K, Unterbeck A, Price DL. Evidence that beta-amyloid protein in Alzheimer's disease is not derived by normal processing. Science 1990; 248: 492-495.
- 28. LaFerla FM, Green KN, Oddo S. Intracellular amyloid-beta in Alzheimer's disease. Nat Rev Neurosci 2007; 8: 499-509.
- 29. Gibbs RB, McCabe JT, Buck CR, Chao MV, Pfaff DW. Expression of NGF receptor in the rat forebrain detected with in situ hybridization and immunohistochemistry. Brain Res Mol Brain Res 1989; 6: 275-287.
- 30. Woolf NJ, Gould E, Butcher LL. Nerve growth factor receptor is associated with cholinergic neurons of the basal forebrain but not the pontomesencephalon. Neuroscience 1989; 30: 143-152.
- 31. Whitehouse PJ, Price DL, Clark AW, Coyle JT, DeLong MR. Alzheimer disease: evidence for selective loss of cholinergic neurons in the nucleus basalis. Ann Neurol 1981; 10: 122-126.

- 32. Mufson EJ, Bothwell M, Kordower JH. Loss of nerve growth factor receptor-containing neurons in Alzheimer's disease: a quantitative analysis across subregions of the basal forebrain. Exp Neurol 1989; 105: 221-232.
- 33. Longo FM, Massa SM. Neurotrophin receptor-based strategies for Alzheimer's disease. Curr Alzheimer Res 2005; 2: 167-169.
- 34. Hu XY, Zhang HY, Qin S, Xu H, Swaab DF, Zhou JN. Increased p75(NTR) expression in hippocampal neurons containing hyperphosphorylated tau in Alzheimer patients. Exp Neurol 2002; 178: 104-111.
- 35. Ginsberg SD, Che S, Wuu J, Counts SE, Mufson EJ. Down regulation of trk but not p75NTR gene expression in single cholinergic basal forebrain neurons mark the progression of Alzheimer's disease. J Neurochem 2006; 97: 475-487.
- 36. Counts SE, Mufson EJ. The role of nerve growth factor receptors in cholinergic basal forebrain degeneration in prodromal Alzheimer disease. J Neuropathol Exp Neurol 2005; 64: 263-272.
- 37. Chakravarthy B, Gaudet C, Menard M, Atkinson T, Brown L, Laferla FM, *et al*. Amyloid-beta peptides stimulate the expression of the p75(NTR) neurotrophin receptor in SHSY5Y human neuroblastoma cells and AD transgenic mice. J Alzheimers Dis 2010; 19: 915-925.
- 38. Yaar M, Zhai S, Fine RE, Eisenhauer PB, Arble BL, Stewart KB, *et al*. Amyloid beta binds trimers as well as monomers of the 75-kDa neurotrophin receptor and activates receptor signaling. J Biol Chem 2002; 277: 7720-7725.
- 39. Smith-Swintosky VL, Pettigrew LC, Craddock SD, Culwell AR, Rydel RE, Mattson MP. Secreted forms of beta-amyloid precursor protein protect against ischemic brain injury. J Neurochem 1994; 63: 781-784.
- 40. Thornton E, Vink R, Blumbergs PC, Van Den Heuvel C. Soluble amyloid precursor protein alpha reduces neuronal injury and improves functional outcome following diffuse traumatic brain injury in rats. Brain Res 2006; 1094: 38-46.
- 41. Corrigan F, Pham CL, Vink R, Blumbergs PC, Masters CL, van den Heuvel C, *et al*. The neuroprotective domains of the amyloid precursor protein, in traumatic brain injury, are located in the two growth factor domains. Brain Res 2011; 1378: 137- 143.
- 42. Ring S, Weyer SW, Kilian SB, Waldron E, Pietrzik CU, Filippov MA, *et al*. The secreted beta-amyloid precursor protein ectodomain APPs alpha is sufficient to rescue the anatomical, behavioral, and electrophysiological abnormalities

of APP-deficient mice. J Neurosci 2007; 27: 7817-7826.

- 43. Moechars D, Lorent K, Dewachter I, Baekelandt V, De Strooper B, Van Leuven F. Transgenic mice expressing an alpha-secretion mutant of the amyloid precursor protein in the brain develop a progressive CNS disorder. Behav Brain Res 1998; 95: 55-64.
- 44. Moechars D, Lorent K, De Strooper B, Dewachter I, Van Leuven F. Expression in brain of amyloid precursor protein mutated in the alpha-secretase site causes disturbed behavior, neuronal degeneration and premature death in transgenic mice. EMBO J 1996; 15: 1265-1274.
- 45. Mattson MP, Cheng B, Culwell AR, Esch FS, Lieberburg I, Rydel RE. Evidence for excitoprotective and intraneuronal calcium-regulating roles for secreted forms of the beta-amyloid precursor protein. Neuron 1993; 10: 243-254.
- 46. Goodman Y, Mattson MP. Secreted forms of beta-amyloid precursor protein protect hippocampal neurons against amyloid beta-peptide-induced oxidative injury. Exp Neurol 1994; 128: 1-12.
- 47. Furukawa K, Sopher BL, Rydel RE, Begley JG, Pham DG, Martin GM, *et al*. Increased activity-regulating and neuroprotective efficacy of alpha-secretase-derived secreted amyloid precursor protein conferred by a C-terminal heparinbinding domain. J Neurochem 1996; 67: 1882-1896.
- 48. Copanaki E, Chang S, Vlachos A, Tschape JA, Muller UC, Kogel D, *et al*. sAPPalpha antagonizes dendritic degeneration and neuron death triggered by proteasomal stress. Mol Cell Neurosci 2010; 44: 386-393.
- 49. Ohsawa I, Hirose Y, Ishiguro M, Imai Y, Ishiura S, Kohsaka S. Expression, purification, and neurotrophic activity of amyloid precursor protein-secreted forms produced by yeast. Biochem Biophys Res Commun 1995; 213: 52-58.
- 50. Ohsawa I, Takamura C, Kohsaka S. The amino-terminal region of amyloid precursor protein is responsible for neurite outgrowth in rat neocortical explant culture. Biochem Biophys Res Commun 1997; 236: 59-65.
- 51. Mahadeo D, Kaplan L, Chao MV, Hempstead BL. High affinity nerve growth factor binding displays a faster rate of association than p140trk binding. Implications for multi-subunit polypeptide receptors. J Biol Chem 1994; 269: 6884-6891.
- 52. Lee R, Kermani P, Teng KK, Hempstead BL. Regulation of cell survival by secreted proneurotrophins. Science 2001; 294: 1945-1948.